

Ca²⁺-activated Cl⁻ currents are dispensable for olfaction

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Canonical olfactory signal transduction involves the activation of cyclic AMP-activated cation channels that depolarize the cilia of receptor neurons and raise intracellular calcium. Calcium then activates Cl⁻ currents that may be up to tenfold larger than cation currents and are believed to powerfully amplify the response. We identified Anoctamin2 (Ano2, also known as TMEM16B) as the ciliary Ca²⁺-activated Cl⁻ channel of olfactory receptor neurons. Ano2 is expressed in the main olfactory epithelium (MOE) and in the vomeronasal organ (VNO), which also expresses the related Ano1 channel. Disruption of *Ano2* in mice virtually abolished Ca²⁺-activated Cl⁻ currents in the MOE and VNO. *Ano2* disruption reduced fluid-phase electro-olfactogram responses by only ~40%, did not change air-phase electro-olfactograms and did not reduce performance in olfactory behavioral tasks. In contrast with the current view, cyclic nucleotide-gated cation channels do not need a boost by Cl⁻ channels to achieve near-physiological levels of olfaction.

Olfaction in mammals involves the binding of odorants to a subset of the large number of different G protein-coupled receptors that are present in the cilia of olfactory sensory neurons (OSNs)¹. These neurons are predominantly found in the MOE, which senses and discriminates myriad volatile compounds. In the mouse, the nose contains additional, apparently more specialized olfactory organs, such the septal organ of Masera and the VNO². Sensory neurons in the VNO are morphologically distinct and use odorant receptors and signal transduction cascades that differ from those found in the MOE.

In the canonical OSN signal transduction pathway, odorant binding to the receptor locally increases cytosolic cAMP through activation of the olfactory G protein G_{olf} and adenylate cyclase type III^{1,2}. cAMP then opens heteromeric cyclic nucleotide-gated (CNG) cation channels³. The resulting influx of Na⁺ and Ca²⁺ depolarizes the plasma membrane and raises the cytosolic Ca²⁺ concentration ([Ca²⁺]_i), thereby activating Ca²⁺-activated Cl⁻ channels^{1,4-10}. All these components of the signal transduction cascade are localized to sensory cilia, which are embedded in the mucus covering the MOE. These cilia provide a large interaction surface for odorants, and the cilia's small diameters facilitate large local increases in cytosolic cAMP and [Ca²⁺]_i.

There has been broad consensus that Ca²⁺-activated Cl⁻ channels powerfully amplify olfactory signal transduction^{1,4-10}. These channels are thought to mediate an outward flow of Cl⁻, which generates a depolarizing current that, in rodents, is five- to tenfold larger than currents through CNG channels^{8,11,12}. A prerequisite for Cl⁻ efflux is an inside-out electrochemical Cl⁻-gradient. It is believed that the cytosolic chloride concentration of OSNs is raised by the Na⁺K⁺2Cl⁻ co-transporter *Nkcc1* (refs. 9,10,13) and that [Cl⁻] is low in the mucus surrounding the cilia^{14,15}. However, mucosal ion concentrations are difficult to measure *in vivo*, and *Nkcc1* knockout mice display attenuated electro-olfactograms (EOGs)^{11,16}, but normal olfactory sensitivity¹⁷.

To investigate the role of Ca²⁺-activated Cl⁻ channels in olfaction, we disrupted *Ano2* in mice. *Ano2* is a member of the *Anoctamin* (*Tmem16*) gene family, which encodes several Ca²⁺-activated Cl⁻ channels¹⁸⁻²⁰.

In agreement with recent results^{13,21-23}, our knockout-controlled immunolabeling showed *Ano2* expression in cilia of OSNs in the MOE, in microvilli of VNO sensory neurons and in synapses of photoreceptors. Additionally, we found *Ano2* in the olfactory bulb. *Ano1* expression overlapped with *Ano2* in the VNO and the retina, but not in the MOE. Patch-clamp analysis showed that Ca²⁺-activated Cl⁻ currents were undetectable in *Ano2*^{-/-} OSNs. Unexpectedly, however, EOGs of *Ano2*^{-/-} mice were reduced by only up to ~40%, and *Ano2*^{-/-} mice were able to smell normally. Our work calls for a revision of the current view that Ca²⁺-activated Cl⁻ channels have a crucial role in mammalian olfaction.

RESULTS

Disruption of *Ano2* and *Ano2* expression pattern

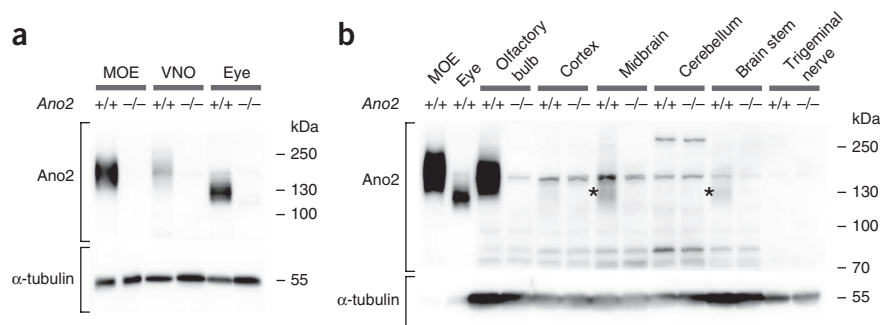
We generated *Ano2*^{-/-} mice by flanking exon 12 with loxP sites and crossing these mice with Cre-recombinase-expressing *deleter* mice²⁴ (Supplementary Fig. 1a,b). The ~160-kDa band that represents the *Ano2* protein in *Ano2*^{+/+} (wild type) olfactory epithelium was absent from *Ano2*^{-/-} tissue (Fig. 1a). Our N-terminal antibody did not detect a truncated protein in *Ano2*^{-/-} tissue, suggesting the occurrence of nonsense-mediated mRNA decay, instability of the truncated protein, or both. The *Ano2*^{-/-} mice were born at a Mendelian ratio; they grew, mated and survived normally and lacked an immediately apparent phenotype.

Consistent with previous data²¹⁻²³, western blots indicated that *Ano2* is most highly expressed in olfactory epithelium (MOE and VNO) and eye tissue (Fig. 1a,b). *Ano2* was expressed at much lower levels in the brain, with the highest expression in the olfactory bulb and signals near the detection limit in the midbrain and brainstem (Fig. 1b). We did not detect expression in the cortex, cerebellum or trigeminal nerve. The differences in apparent sizes of *Ano2* among various tissues are due largely to a higher degree of glycosylation in olfactory tissues (Supplementary Fig. 2). Immunolabeling showed that *Ano2* is expressed on the apical surface of both the MOE and the VNO (Fig. 2a). In the MOE, it colocalized with the cilia marker

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Figure 1 Immunoblot analysis of An $o2$ expression. (a) An $o2$ expression in the MOE, VNO and eye from An $o2^{+/+}$, but not An $o2^{-/-}$, mice. Protein loaded: 16 μ g (MOE) and 100 μ g (other lanes). (b) An $o2$ is detected in lower amounts in the olfactory bulb, midbrain and brainstem, but not in the cortex, cerebellum and trigeminus. The loading control was α -tubulin. Protein loaded: MOE, 1.6 μ g; eye, 10 μ g; other lanes, 50 μ g. The sharp ~140-kDa band remaining in An $o2^{-/-}$ tissue is nonspecific. The broad band (*) in midbrain and brain stem tissue represents An $o2$, as it is absent from An $o2^{-/-}$ tissue and runs at a similar size as in the retina.



protein acetylated tubulin (Fig. 2b) and the Cnga2 subunit of the olfactory CNG channel^{3,25,26} (Fig. 2c). An $o2$ staining was specific, as we observed no signal in An $o2^{-/-}$ tissue (Fig. 2c).

We also detected An $o2$ in vomeronasal sensory neurons (VSNs), where it colocalized with the closely related An $o1$ Cl⁻ channel in sensory protrusions (Fig. 3a). In the MOE, however, we did not detect An $o1$ in olfactory sensory neurons (Fig. 3b,c), but it was present in the Bowman's glands (not shown). Similarly, An $o1$ was present in the apical membranes of the nasal glands (Fig. 3b,d) and in the non-ciliated cells of the respiratory epithelium (Fig. 3b,c), consistent with a role in salt secretion, as in other epithelia^{18,27,28}.

An $o2$ protein was present on the OSN axons that converge on the glomeruli in the olfactory bulb (Fig. 4a,b). In the eye, our knockout-controlled staining confirmed the presence of An $o2$ in the outer plexiform layer of the retina²² (Supplementary Fig. 3a). An $o2$ colocalized with the PSD-95 adaptor protein (Supplementary Fig. 3b) and the Ca²⁺-ATPase PMCA (Supplementary Fig. 3c) in the presynaptic structures of photoreceptors, where these proteins form a complex together with MPP4 (ref. 22). Although An $o2$ expression is lost from the photoreceptors of Mpp4^{-/-} mice²², loss of An $o2$ affected neither expression of PSD-95 nor PMCA (Supplementary Fig. 3b,c). An $o1$ antibodies labeled the same presynaptic structures that we had also identified as An $o2$ positive (Supplementary Fig. 3a).

No change of key proteins and axonal convergence

Neither immunohistochemistry (Fig. 3a) nor immunoblotting (Supplementary Fig. 4a) indicated an upregulation of An $o1$ in

the VNO or MOE of An $o2^{-/-}$ mice. Consistent with the normal morphology of both olfactory organs in An $o2^{-/-}$ mice (Figs. 2c and 3a), expression levels of the olfactory marker protein OMP²⁹ were unchanged (Supplementary Fig. 4b). An $o2$ disruption caused no change in expression of the downstream target of G_{olf} adenylate cyclase III (Supplementary Fig. 4c,f), or the cAMP-activated cation channel subunit Cnga2 (Fig. 2c and Supplementary Fig. 4d). Expression of Nkcc1, An $o6$, An $o8$ and An $o10$ in the MOE was also unaltered (Supplementary Fig. 4f).

Immunohistochemistry (Fig. 4a) and immunoblotting (Supplementary Fig. 4e) showed no change in the amount of tyrosine hydroxylase in the olfactory bulb, either. The expression of this enzyme depends on neuronal activity and is severely reduced in the olfactory bulb of anosmic Cnga2^{-/-} mice³⁰. OSN axon coalescence on glomeruli is perturbed in mice lacking functional CNG channels³¹ or the adenylate cyclase³². We detected no obvious change in axonal convergence for OSNs expressing either the P2 or M72 receptor³¹ (Fig. 4c-e). These observations suggest that olfaction is not grossly impaired in An $o2^{-/-}$ mice and that intrinsic OSN activity is not changed to a degree that changes the olfactory map.

Ca²⁺-activated Cl⁻ currents are absent from An $o2^{-/-}$ OSNs

Whole-cell patch-clamp analysis of OSNs from the MOE *in situ* under conditions that largely suppress cation currents (Fig. 5) revealed Ca²⁺-activated Cl⁻ currents (Fig. 5d,f,g,i) that were not observed in the absence of intracellular Ca²⁺ (Fig. 5a-c). These currents showed typical time dependence and outward rectification at 1.5 μ M free Ca²⁺ (Fig. 5d,f), but not at 13 μ M free Ca²⁺ (Fig. 5g,i), similar to data reported for Ca²⁺-activated Cl⁻ currents for heterologously expressed An $o1$ and An $o2$ (refs. 20–22,33,34) and for

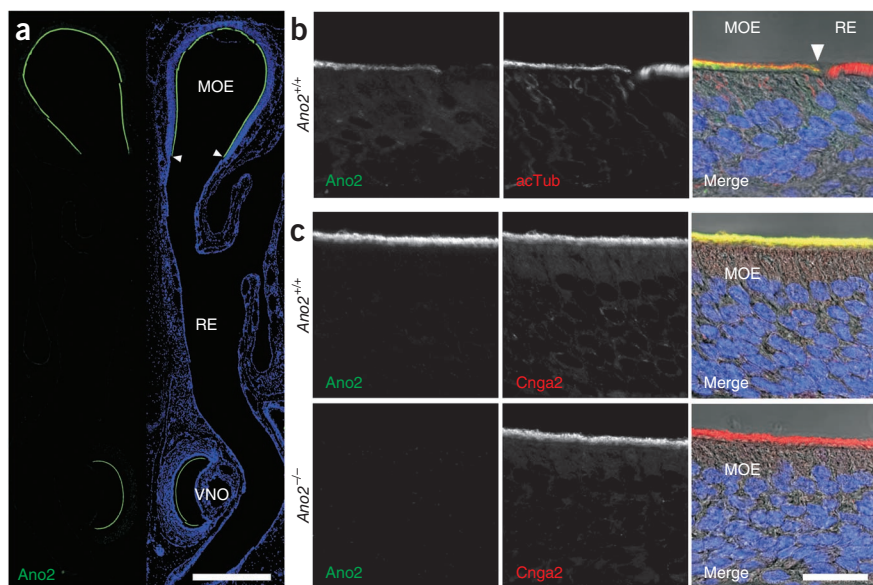


Figure 2 An $o2$ localizes to sensory protrusions in olfactory epithelia. (a) Coronal view of the anterior part of the nasal cavity stained for An $o2$ (antibody rbAn $o2$ _N3-3). An $o2$ localized to the surface of the MOE and the VNO, but not to the respiratory epithelium (RE). Nuclei (blue) are shown only in the right half of the picture. Scale bar, 500 μ m. (b) Confocal images of the transition region from MOE to respiratory epithelium shows An $o2$ expression exclusively in OSNs, where it colocalizes with acetylated tubulin (acTub). (c) Colocalization of An $o2$ and Cnga2 in the MOE. Merged images with nuclei labeled in blue are superimposed on transmission light pictures. Scale bar, 20 μ m. Arrowheads mark the transition from MOE to respiratory epithelium.

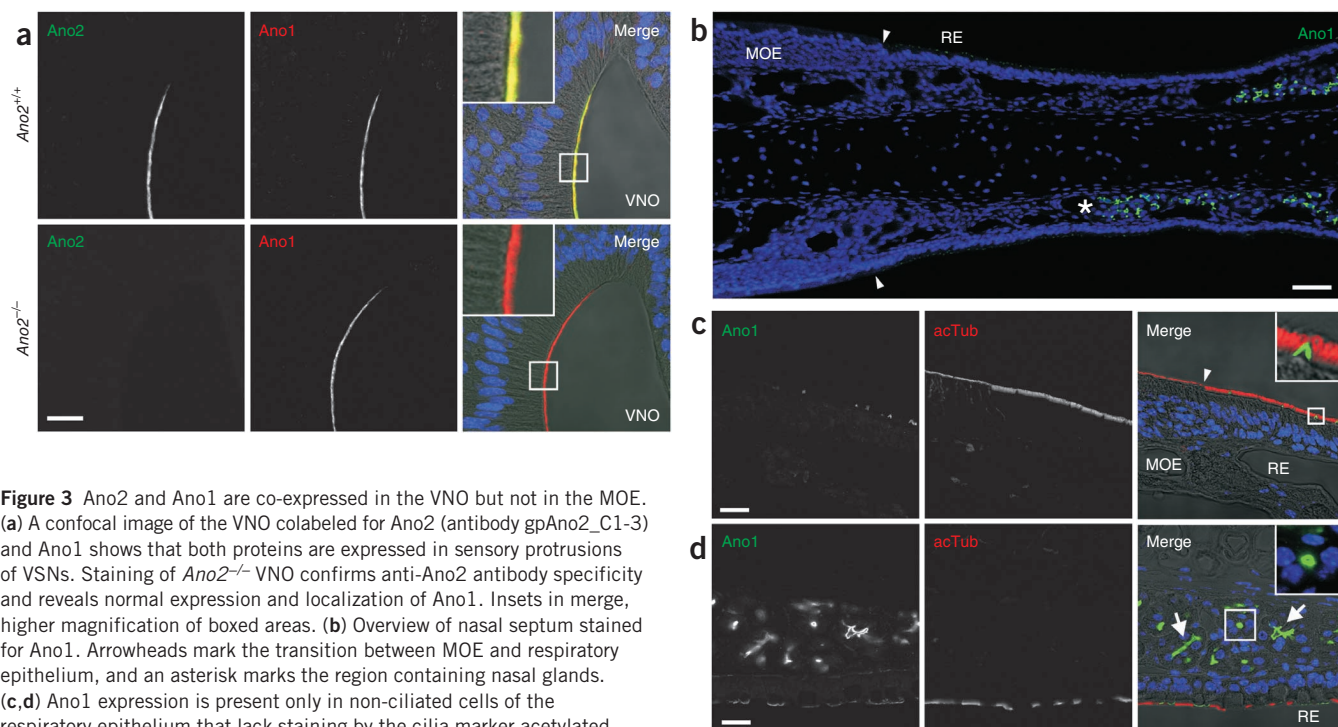


Figure 3 *Ano2* and *Ano1* are co-expressed in the VNO but not in the MOE. (a) A confocal image of the VNO colabeled for *Ano2* (antibody gp*Ano2_C1-3*) and *Ano1* shows that both proteins are expressed in sensory protrusions of VSNS. Staining of *Ano2*^{-/-} VNO confirms anti-*Ano2* antibody specificity and reveals normal expression and localization of *Ano1*. Insets in merge, higher magnification of boxed areas. (b) Overview of nasal septum stained for *Ano1*. Arrowheads mark the transition between MOE and respiratory epithelium, and an asterisk marks the region containing nasal glands. (c,d) *Ano1* expression is present only in non-ciliated cells of the respiratory epithelium that lack staining by the cilia marker acetylated tubulin (acTub) (highlighted in high-magnification insets in c). Beneath the respiratory epithelium, *Ano1* is found in the apical membranes of nasal gland epithelia (arrows and high-magnification inset in d). Nuclei are labeled in blue, and merged images are superimposed on transmission light pictures. Scale bars, 20 μ m (a,c-d), 50 μ m (b).

frog OSNs⁷. We could not detect these Cl⁻ currents in *Ano2*^{-/-} OSNs (Fig. 5e,f,h,i). To exclude the possibility that transient Ca²⁺-activated Cl⁻ currents remain in *Ano2*^{-/-} OSNs, we used flash photolysis of

caged Ca²⁺ and 8-Br-cAMP with isolated receptor neurons (Fig. 6). As observed previously^{12,35}, *Ano2*^{+/+} neurons responded to uncaging of Ca²⁺ with large, rapidly activating, transient currents that

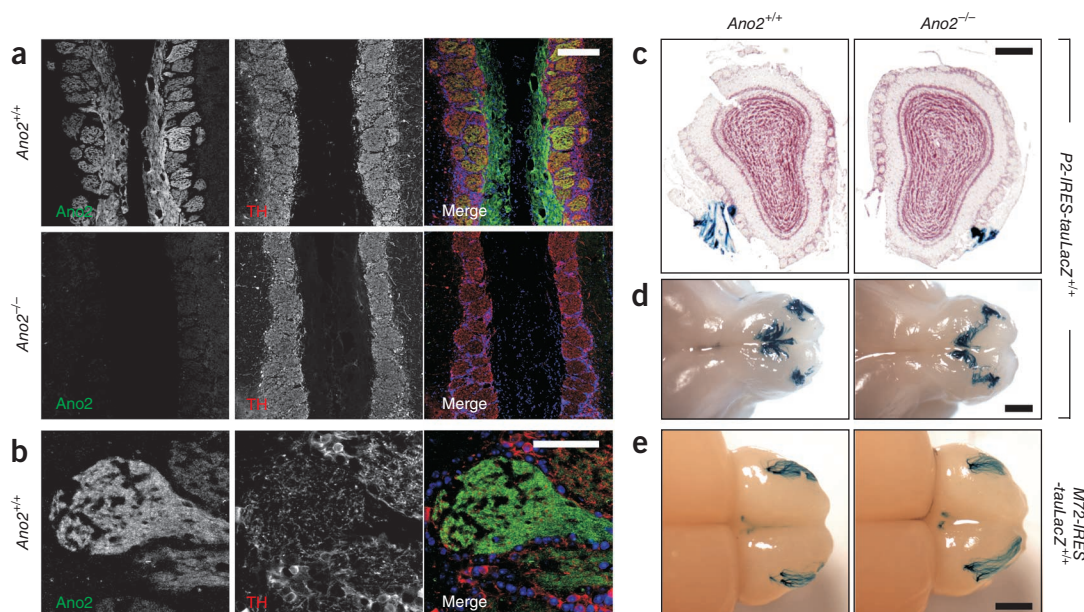


Figure 4 The olfactory bulb in *Ano2*^{-/-} and *Ano2*^{+/+} mice. (a) Specific staining for *Ano2* in the olfactory nerve layer and in axons entering olfactory glomeruli. The section shows part of the olfactory bulb from both hemispheres, with the cleft in center. Activity-dependent tyrosine hydroxylase (TH) expression in periglomerular cells is not changed in the *Ano2*^{-/-} section. (b) Higher magnification of a glomerulus stained for *Ano2* and TH. Scale bars: 200 μ m (a) and 50 μ m (b). (c-e) Axonal coalescence of OSNs expressing the P2 (c,d) or M72 (e) olfactory receptors on glomeruli in the olfactory bulb. Olfactory bulbs of homozygous P2-IRES-tauLacZ or M72-IRES-tauLacZ mice³¹ with disrupted or wild-type *Ano2* genes were stained for LacZ. Bulbi were sectioned (c), or the ventral (d) or dorsal (e) side of intact bulbi was observed. No difference in coalescence was seen (number of mouse pairs: P2, $n = 9$ (4–15 weeks old), M72, $n = 2$ (5 weeks old) and $n = 1$ (24 weeks old)). Scale bars, 0.5 mm (c), 1 mm (d,e).

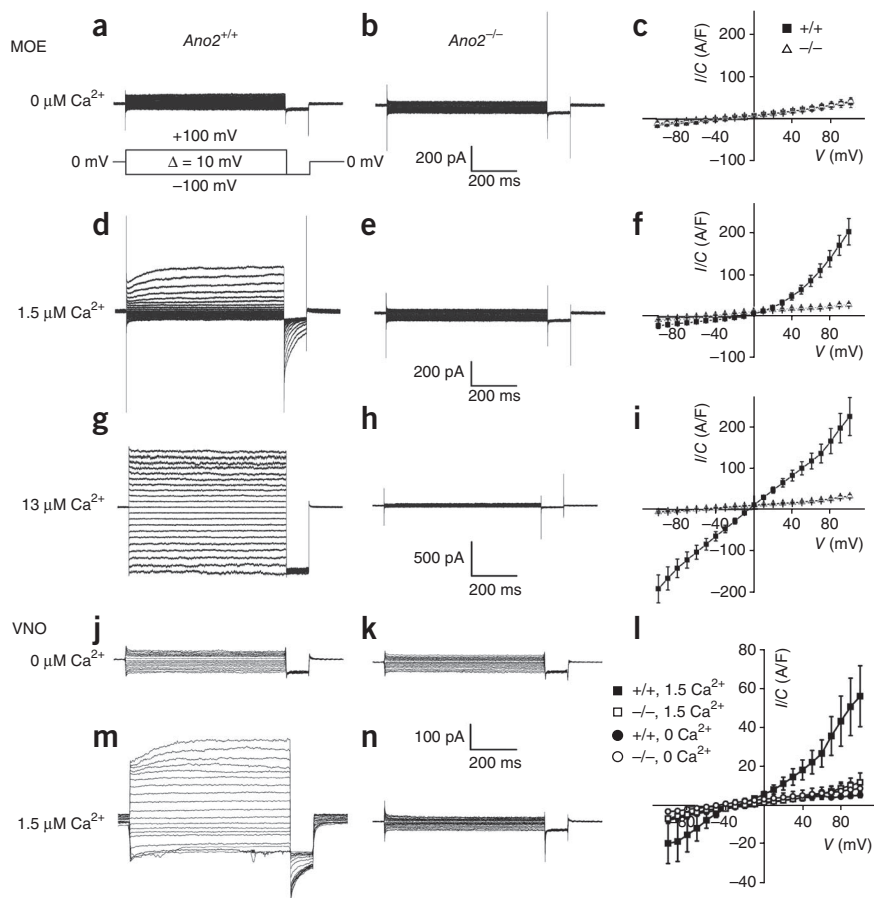


Figure 5 Effect of *Ano2* disruption on Ca^{2+} -activated Cl^- currents. Patch-clamp recordings of olfactory receptor neurons from the MOE (a–i) and the VNO (j–l). (a,d,g) Typical current traces obtained from *Ano2*^{+/+} OSNs in the presence of nominally 0 μM , 1.5 μM and 13 μM Ca^{2+} in the recording pipette, respectively. The voltage-clamp protocol is shown in a. (b,e,h) Current densities (I/C) from *Ano2*^{-/-} OSNs under conditions as in a,d,g. (c,f,i) Averaged current-voltage relationships of steady-state currents with 0 μM , 1.5 μM and 13 μM Ca^{2+} in the pipette, respectively. ■ wild-type cells, △ *Ano2*^{-/-} cells. Error bars, s.e.m. Number of cells measured: (c) 7 wild type, 7 knockout; (f), 15 wild type, 11 knockout; (i), 14 wild type, 10 knockout. (j,m) Typical current traces of wild-type vomeronasal sensory neurons (VSNs) with 0 μM Ca^{2+} (j) and 1.5 μM free Ca^{2+} (m) in the pipette. (k) *Ano2*^{-/-} VSN with 0 μM Ca^{2+} , and (n) with 1.5 μM Ca^{2+} in the pipette. (l) Averaged current-voltage relationships from VNO receptor neurons measured with 1.5 μM Ca^{2+} (■ *Ano2*^{+/+}, □ *Ano2*^{-/-}; n = 7 and 6, respectively) and with 0 μM Ca^{2+} (● wild type, ○ *Ano2*^{-/-}; n = 5 for both). Error bars, s.e.m. Voltage clamp protocol as in a.

larger. Averaged current/voltage curves revealed that Ca^{2+} -activated Cl^- currents of VSNs depend predominantly on *Ano2* (Fig. 5l). Although *Ano1* is expressed in the VNO (Fig. 3a), its contribution to VSN currents seems minor.

reversed close to the Cl^- equilibrium potential (Fig. 6a,c). Currents in *Ano2*^{-/-} OSNs were reduced to less than 10% of wild-type currents and reversed close to the K^+ equilibrium potential. Flash release of 8-Br-cAMP elicited large currents in *Ano2*^{+/+} but not *Ano2*^{-/-} OSNs (Fig. 6b). Currents remaining in *Ano2*^{-/-} neurons probably represent cation currents through CNG channels¹². We conclude that Ca^{2+} -activated Cl^- currents of OSNs are mediated by *Ano2*.

In the VNO, both *Ano2*^{+/+} and *Ano2*^{-/-} VSNs had low background currents in the absence of intracellular Ca^{2+} (Fig. 5j–l). In *Ano2*^{+/+} VSNs, 1.5 μM [Ca^{2+}]_i elicited Cl^- currents with an outwardly rectifying, time-dependent component (Fig. 5l,m). Currents of most *Ano2*^{-/-} VSNs were indistinguishable from those we observed without Ca^{2+} (Fig. 5n), but a few cells showed currents up to twofold

Disruption of *Ano2* moderately reduces EOGs

We investigated the role of *Ano2* in the MOE response to odorants using EOGs *ex vivo* (Fig. 7). In fluid-phase EOGs, we continuously superfused the surface of turbinates with Ringer solution and applied an odorant cocktail (Fig. 7a,b). The superfusate could be switched to a solution containing niflumic acid (NFA). NFA has been widely used to block Ca^{2+} -activated Cl^- channels in OSNs and other cells^{4,6,9,11,16,34,36,37} and inhibits both *Ano1* and *Ano2* (refs. 18,20,21,33,34). The response to odorants was reduced by roughly 40% in *Ano2*^{-/-} EOGs as compared to the wild type (Fig. 7e). We observed similar reductions in EOG amplitudes caused by *Ano2* disruption when adenylyl cyclase was directly activated by forskolin (Fig. 7c–e), or with single odorants (Fig. 7f). As expected, NFA-insensitive voltage

Figure 6 Response of isolated olfactory receptor neurons to photoreleased Ca^{2+} or 8-Br-cAMP.

(a) Typical current responses to a sudden [Ca^{2+}] increase in isolated OSNs from both genotypes in whole-cell patch-clamp experiments under symmetrical Cl^- . Cells were clamped to -50 mV, 0 mV and +50 mV. Arrows indicate the flash that releases Ca^{2+} from DMNP-EDTA. Inset, larger magnification to reveal the currents remaining in *Ano2*^{-/-} OSNs. (b) Typical currents elicited by flash release of 8-Br-cAMP (arrows) in isolated OSNs from *Ano2*^{+/+} and *Ano2*^{-/-} mice held at -50 mV. Mean amplitudes were -445 ± 111 pA and -43.2 ± 9.9 pA for *Ano2*^{+/+} and *Ano2*^{-/-} OSNs, respectively (\pm s.e.m., $P = 0.0016$). (c) Peak currents elicited by uncaging Ca^{2+} as function of holding voltage, averaged from eight *Ano2*^{+/+} and seven *Ano2*^{-/-} OSNs with three or more mice per genotype. Error bars, s.e.m. Significance levels (two sample t-test): *, $P < 0.05$; **, $P < 0.01$.

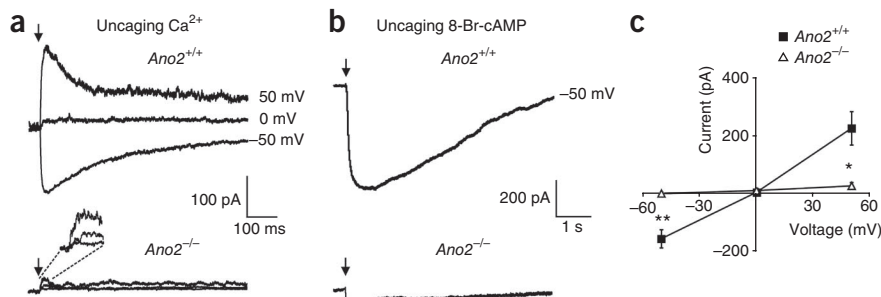
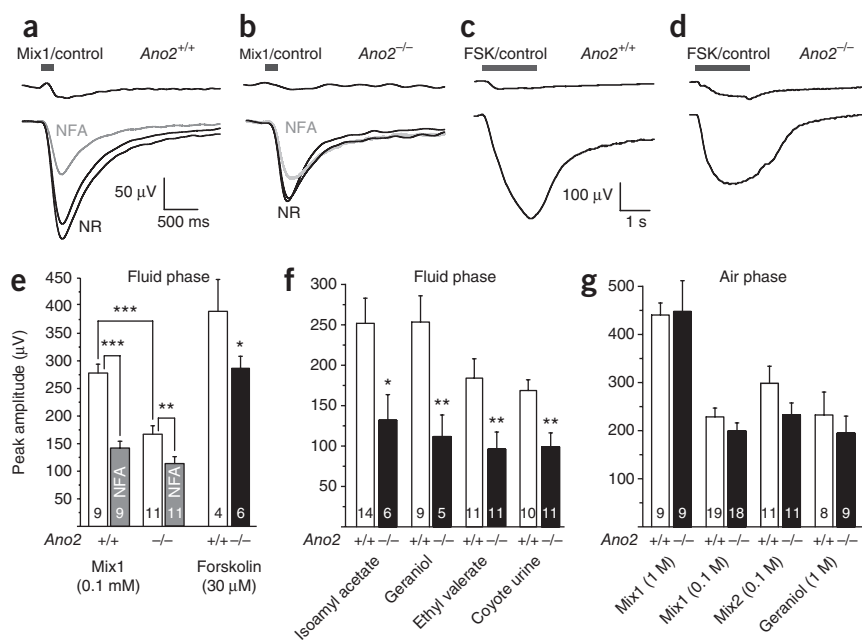


Figure 7 EOGs from the MOE. (a,b) Typical fluid-phase EOGs from *Ano2*^{+/+} (a) and from *Ano2*^{-/-} (b) mice with a mixture of 17 odorants ('Mix1': isopentyl acetate, hexanal, eucalyptol, limonene, 2-heptanone, menthyl acetate, peppermint oil, eugenol, ethyl valerate, ethyl butyrate, ethyl tiglate, allyl tiglate, octanal, isobutyl propionate, acetal, hexanoic acid, 2-hexanone; 100 μ M each). Bars above traces indicate application (200 ms) of odorant mixture (lower traces) or vehicle (upper trace). Gray traces were obtained with 300 μ M NFA and black traces in normal Ringer (NR) before and after exposure to NFA (washout). Traces were averaged from responses to ten repeated stimuli. (c,d) Typical fluid-phase EOGs evoked by 2-s application of 30 μ M forskolin (lower trace) or vehicle control (upper trace) to *Ano2*^{+/+} (c) and *Ano2*^{-/-} MOE (d).



(e) Averaged EOG amplitudes from experiments as in a-d. (f) Averaged fluid-phase EOG amplitudes elicited by individual odorants at 1 mM concentration and with 1:20 diluted coyote urine. (g) Averaged air-phase EOG amplitudes from mice of both genotypes. 'Mix1' is the same odorant mixture as in a.

'Mix2' contained eight odorants (d-carvone, 1-heptanol, 2-methylbutyric acid, geraniol, isopentylamine, 2-hexanone, acetophenone, 1-octanal). Differences in responses between genotypes are not statistically significant. Numbers in columns, number of measurements; error bars, s.e.m. Significance levels (two sample *t*-test): *, $P < 0.05$; **, $P < 0.01$; ***, $P < 0.001$.

excursions were similar in both genotypes (Fig. 7e). Notably, NFA also inhibited odorant-induced responses in *Ano2*^{-/-} epithelia (Fig. 7b,e). Comparison between *Ano2*^{+/+} and *Ano2*^{-/-} mice revealed that only ~60% of the response to NFA was due to an inhibition of *Ano2*. Hence, previous studies^{9,11,37} may have overestimated the role of Ca^{2+} -activated Cl^{-} channels in olfaction.

In air-phase EOGs, we exposed the surfaces of turbinates to water-saturated air, and we applied an odorant or vehicle using an air puff. In this configuration, mucosal ion concentrations are expected to more closely match physiological levels. In contrast with our results for fluid-phase EOGs (Fig. 7a-f), we detected no significant differences between the genotypes in air phase (Fig. 7g).

With appropriate driving forces, apical Cl^{-} channels may change mucosal ion composition, similar to the proposed role of *Ano1* in

secretory epithelia^{18,27,28}. However, Cl^{-} -sensitive microelectrode measurements showed no difference in mucosal $[Cl^{-}]$ between the genotypes (*Ano2*^{+/+}: 84.0 ± 11.8 mM (s.e.m., $n = 6$); *Ano2*^{-/-}: 84.4 ± 9 mM (s.e.m., $n = 11$)). These values agree with data from amphibia¹⁵.

No olfactory deficits detected in *Ano2*^{-/-} mice

Using an automated olfactometer^{17,30,38,39}, we tested the mice using an associative olfactory learning task. The mice were trained to sample two different stimuli, one of which was associated with a water reward, and to lick in response to only the rewarded odor.

Disruption of *Ano2* did not affect the mice's performance in discrimination tests in which they learned to distinguish geraniol from the mineral oil solvent (Fig. 8a). By contrast to *Ano2*^{-/-} mice, *Cnga2*^{-/-} mice³¹ showed a clear-cut impairment in the

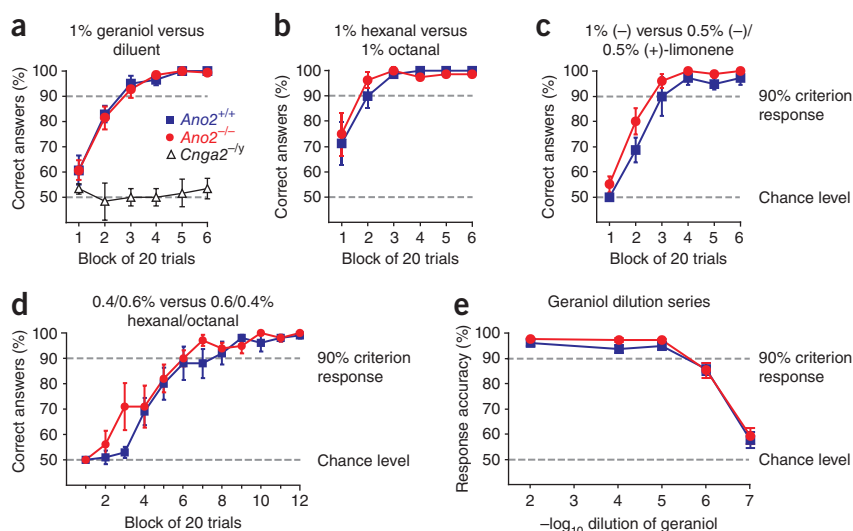


Figure 8 *Ano2* disruption affected neither odor discrimination nor olfactory sensitivity. (a-d) Different discrimination tasks. As true for wild-type littermates, *Ano2*^{-/-} mice learned to discriminate between 1% geraniol and the diluent mineral oil ($n = 6$ for each genotype) (a), between 1% hexanal and 1% octanal ($n = 3$ for each genotype) (b), between 1% (-)-limonene and an enantiomeric mixture of 0.5% (-)-limonene and 0.5% (+)-limonene ($n = 4$ for each genotype) (c) and between 0.4% hexanal/0.6% octanal and 0.6% hexanal/0.4% octanal ($n = 5$ for each genotype) (d). Anosmic *Cnga2*^{-/-} mice ($n = 3$) could not detect 1% geraniol (a). (e) Odor detection threshold for geraniol. Both *Ano2*^{-/-} ($n = 6$) and *Ano2*^{+/+} littermates ($n = 6$) detected geraniol and discriminated it from the diluent only down to a dilution of 10^{-6} . The first data point with a geraniol dilution of 10^{-2} corresponds to a. Error bars, s.e.m. There was no significant difference between *Ano2*^{+/+} and *Ano2*^{-/-} mice in any of these tests.

same test (Fig. 8a), as reported^{25,30}. Discrimination ability was neither reduced in more complex discrimination tasks such as distinguishing hexanal from octanal (Fig. 8b), or differentiating between (–)-limonene and an enantiomeric mixture of equal parts of (+)- and (–)-limonene (Fig. 8c). Discrimination between 0.4% hexanal (vol/vol) / 0.6% octanal (vol/vol) and 0.6% hexanal (vol/vol) / 0.4% octanal (vol/vol) was even more difficult, as evident from the larger number of trials needed to distinguish both mixtures (Fig. 8d). Even in this task, *Ano2*^{–/–} mice performed similarly to wild type. Using successive dilutions of geraniol, we tested whether the loss of *Ano2* affected odor sensitivity. With either genotype, odor detection dropped sharply when the dilution reached 10^{–7} (Fig. 8e). We conclude that Ca²⁺-activated Cl[–] channels are not essential for olfaction.

DISCUSSION

Ca²⁺-activated Cl[–] channels are prominently expressed in olfactory sensory neurons. In freshwater animals, in which ion gradients preclude a depolarizing influx of monovalent cations, Cl[–] efflux may generate almost all of the receptor current^{5,40}. By contrast, the cation concentration in the mucus surrounding olfactory cilia in mammals^{14,15} allows for a sizeable depolarization when CNG channels open upon an odorant-induced rise in cAMP. After first publications had shown that Ca²⁺-activated Cl[–] currents could powerfully amplify olfactory receptor currents^{4–7}, this concept has received support from many groups. Experiments with isolated receptor neurons^{4–6,8} showed that up to 80–90% of the receptor current is carried by Cl[–]. An important role for Cl[–] in olfaction was also inferred from EOGs^{11,16,37}. However, as the molecular identity of the olfactory Ca²⁺-activated Cl[–] channel has remained elusive³⁵, the role of Cl[–] channels in olfaction could not be tested directly.

Olfactory cilia express Bestrophin-2 (*Best2*; ref. 41) and *Ano2* (refs. 13,21), which were both hypothesized to mediate ciliary Ca²⁺-activated Cl[–] currents. A role of Bestrophin-2 has recently been excluded by normal Cl[–] currents in *Best2*^{–/–} OSNs³⁵. The present work unambiguously identifies *Ano2* as the olfactory Ca²⁺-activated Cl[–] channel, or at least as an essential component thereof. Although *Ano1* is expressed in the VNO, its contribution to Ca²⁺-activated Cl[–] currents in VSNs seems minor.

Unexpectedly, *Ano2*^{–/–} mice had no detectable impairment of olfaction. This is not due to a compensatory upregulation of other Ca²⁺-activated Cl[–] channels or of the cyclic nucleotide-gated channel. The normal growth and survival of newborn *Ano2*^{–/–} mice indicated that they could smell the teats of their mothers. Near-normal olfaction was suggested indirectly by unaltered tyrosine hydroxylase expression in the olfactory bulb and by apparently normal axonal coalescence on glomeruli in the olfactory bulb, and it was confirmed by quantitative olfactometry. In contrast to *Ano2*^{–/–} mice, *Cnga2*^{–/–} mice have severely impaired olfaction (refs. 25,30 and present work). We conclude that CNG-mediated cation currents do not need an additional boost from anion currents for near-physiological levels of olfactory sensitivity.

Although fluid-phase EOGs revealed a significant difference between the genotypes, the ~40% reduction of *Ano2*^{–/–} EOG amplitudes is much less than expected from the published 80–90% contribution of Cl[–] to OSN receptor currents^{8,11,12}. The contribution of Ca²⁺-activated Cl[–] channels was often estimated from the inhibition by NFA^{11,37}, but this compound also modulates several other ion channels (for example, refs. 42,43). The attenuation of *Ano2*^{–/–} EOGs by NFA now demonstrates that NFA also affects MOE targets other than Ca²⁺-activated Cl[–] channels. Hence, previous studies based on NFA inhibition^{11,37} have overestimated the role

of Ca²⁺-activated Cl[–] channels. These channels might contribute only about 40% to receptor currents *in vivo*.

The ~40% reduction of fluid-phase EOG in *Ano2*^{–/–} mice fits well with the ~39% to ~57% reduction of EOG amplitudes in *Nkcc1*^{–/–} mice^{11,16}. The Na⁺K⁺2Cl[–] co-transporter *Nkcc1* may accumulate Cl[–] into OSNs^{9–11,13}, a prerequisite for depolarizing Cl[–] currents. On the basis of the NFA-mediated block of *Nkcc1*^{–/–} EOG responses, the authors¹¹ concluded that a substantial depolarizing Cl[–] current remained in *Nkcc1*^{–/–} OSNs and proposed that other transporters contribute to intracellular Cl[–] accumulation^{11,16}. The sizeable non-specific component of NFA inhibition observed here, however, suggests that their results^{11,16} are consistent with *Nkcc1* being the main Cl[–] accumulator of OSNs. In *Nkcc1*^{–/–} mice, intraciliary [Cl[–]] is then expected to be close to equilibrium, and opening of *Ano2* channels should shunt rather than amplify receptor currents. Such a shunting inhibition may explain the larger decrease of EOG amplitudes in *Nkcc1*^{–/–} mice compared to *Ano2*^{–/–} mice.

How can the roughly 40% reduction of fluid-phase EOG amplitude, and almost no reduction of air-phase EOGs, be reconciled with previous studies that reported a large, up to 80–90% contribution of Cl[–] to receptor currents? Many of those studies used isolated receptor neurons that were patch-clamped at the cell body^{1,4–8,12}. Also, our experiments on isolated OSNs suggest an ~90% contribution of *Ano2* to cAMP-induced currents. The disruption of tight junctions during isolation of OSNs may allow ciliary *Ano2* channels to spread to the cell body, where their electrical accessibility is much better¹⁰. The length (20–30 μm) and small diameter (0.1 μm) of cilia imply that channels in their distal part contribute little to voltage changes at the cell body, in particular when ciliary membrane conductance is high during signal transduction⁴⁴.

Another important consequence of ciliary geometry is the rapid dissipation of ion gradients. Taking into account ion concentrations that were measured in the mucus and the dendritic knob (as a surrogate for cilia)^{10,14,15}, Cl[–] gradients across the ciliary membrane should dissipate more easily than do those for Na⁺ or Ca²⁺, which carry depolarizing currents through CNG channels. *Nkcc1* is thought to rapidly replenish ciliary Cl[–], which exits through anion channels during signal transduction^{10,13}. Under physiological conditions, the driving force for *Nkcc1*-mediated ciliary Cl[–] uptake is small¹⁰ owing to rather low mucosal [Na⁺] (55 mM or 85 mM^{14,15}) and [Cl[–]] (55 mM or 93 mM^{14,15}; 84 mM (this work)). The driving force for Cl[–] uptake is larger in extracellular Ringer-type solutions (>140 mM NaCl) that were used with isolated OSNs. Hence, more efficient ciliary Cl[–] replenishment from Ringer solution than from mucus may lead to larger signal amplification by Cl[–] channels in isolated OSNs and in fluid-phase EOGs than are observed in air-phase EOGs and olfaction *in vivo*.

If the 40% reduction in *Ano2*^{–/–} fluid-phase EOG amplitude translates into a similar reduction of action potential firing *in vivo*, our olfactory tests may not be sensitive enough to detect ensuing moderate changes in olfaction thresholds. On the other hand, air-phase EOGs, which more closely reflect the physiological situation, were not substantially impaired by *Ano2* disruption, suggesting that the olfactory information that is conveyed to the brain might be nearly unchanged. Notably, there are patients with von Willebrand disease who carry a large genomic deletion⁴⁵ that includes *ANO2* (*TMEM16B*). Consistent with the present work, no impairment of olfaction was reported. The expression of Ca²⁺-activated Cl[–] channels in mammalian OSNs may be an evolutionary vestige from freshwater animals^{5,40}, or it may serve also in mammals to allow consistent olfaction in face of variations of extracellular ion concentrations. *Ano2* may also affect the mucosal fluid film in the face of strong odors, although we showed an

unaltered mucosal Cl⁻ concentration under steady-state conditions. Alternatively, Ano2 might confer a slight increase in olfactory sensitivity that provides an evolutionary advantage. Clearly, however, Ca²⁺-activated Cl⁻ channels are dispensable for near-normal olfaction, and the importance of these channels for olfactory signal amplification has been overestimated.

METHODS

Methods and any associated references are available in the online version of the paper at <http://www.nature.com/natureneuroscience/>.

Note: Supplementary information is available on the Nature Neuroscience website.

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AUTHOR CONTRIBUTIONS

G.M.B. generated *Ano2*^{-/-} mice and Ano2 antibodies, designed, performed and evaluated expression analysis and immunohistochemistry, olfactometry and mouse behavioral studies and wrote the paper. B.P. designed, performed and evaluated patch-clamp analysis, EOG and ion-selective microelectrode measurements and wrote the paper. B.P. and P.F. designed, performed and evaluated flash-photolysis experiments. T.J.J. initiated the project, designed and evaluated experiments and wrote the paper.

COMPETING FINANCIAL INTERESTS

The authors declare no competing financial interests.

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ONLINE METHODS

Ano2^{-/-} mice. An 11-kb genomic region of *Ano2* spanning exons 12 and 13 was cloned from R1 embryonic stem cells by PCR. Exon 12 was flanked with *loxP* sites, a FRT-flanked neomycin selection cassette inserted (**Supplementary Fig. 1**) and the construct cloned into pKO Scrambler plasmid 901 (Lexicon Genetics). The linearized targeting vector was electroporated into R1 embryonic stem cells (129/SvJ). Cells from two independent correctly targeted clones were injected into C57BL/6 blastocysts. Chimeras were crossed to C57BL/6 Cre *deleter* mice²⁴ for *in vivo* excision of exon 12, which eliminates part of the first extracellular loop and part of the second transmembrane domain. Splicing from exon 11 to exon 13 introduces a frameshift after the first transmembrane domain. Mice were in a mixed C57BL/6-129/SvJ background and littermates served as controls. The mice in experiments were older than 4 weeks. We carried out genotyping PCR with primers flanking the *loxP* site of the *Ano2^{-/-}* allele (5'-GGACACCCCGTACTTGAAGA-3' and 5'-AGCACAATGCAGACCAAGTT-3'), yielding 163-bp and 1,051-bp products for the *Ano^{-/-}* and *Ano^{+/+}* alleles, respectively.

Mouse husbandry. Animal care and experiments conformed to German animal protection laws and were approved by the Berlin authorities (LaGeSo). *Cnga2^{-/-}* (previously called *Ocncl^{-/-}*), P2-IRES-tauLacZ and M72-IRES-tauLacZ mice were provided by P. Mombaerts (Max Planck Institute of Biophysics, Frankfurt) and have been described³¹.

Generation of Ano2 antibodies. Antibody to the N-terminal epitope (¹²⁸NGETGKERHGGGPGDVELG) of Ano2 (rbAno2_N3-3) was raised in rabbits; antibody to the extreme C terminus (⁹⁷⁰GDRSRRAASSAPSGRSQP) of Ano2 (gpAno2_C1-3) was raised in guinea pigs (Pineda Antikörper-Service). Peptides were coupled to keyhole limpet hemocyanine by C- and N-terminally added cysteines, respectively. The animals were immunized seven times, and the antibodies were affinity-purified from final bleeds and tested using *Ano2^{-/-}* tissue as a control.

Immunohistochemistry. Anesthetized mice were perfused with PBS followed by 4% paraformaldehyde (PFA) (wt/vol) in PBS. Tissues were post-fixed for 2 h (olfactory bulb) or overnight (nose, eye). We cut 6- μ m cryosections from the eyes and 10- μ m cryosections from the olfactory bulb. Noses were decalcified in 10% EDTA (wt/vol) in PBS for 7 d and post-fixed for 2 h, and then 6- μ m paraffin sections were prepared. For antigen retrieval, sections were boiled for 5 min in 10 mM citrate buffer (pH 6.0) or 10 mM Tris, 1 mM EDTA (pH 9.0), and transferred to 3% BSA (wt/vol) in TBS with 0.2% NP-40 (vol/vol) for blocking and antibody incubation. Antibodies were used at the following dilutions: rbAno2_N3-3 (1:1,000); gpAno2_C1-3 (1:250); anti-TMEM16A (Ano1) rabbit polyclonal (1:100, ab53212, Abcam); its specificity had been shown using *Ano1^{-/-}* tissue⁴⁶); anti-Cnga2 goat polyclonal (1:50, CNG-2/M-20, sc-13700, Santa Cruz); anti-acetylated tubulin mouse monoclonal (1:1,000, clone 6-11B-1, T7451, Sigma-Aldrich); anti-tyrosine hydroxylase mouse monoclonal (1:1,000, MAB5280, Chemicon); anti-PSD-95 mouse monoclonal (1:250, clone 7E3-1B8, MA1-046, ThermoScientific); anti-PMCA mouse monoclonal (1:500, clone 5F10, MA3-914, Affinity BioReagents). Detection was carried out using secondary antibodies coupled to Alexa fluorophores (Invitrogen). Nuclei were stained with Hoechst 33258. Pictures were taken with a Zeiss LSM 510 META laser-scanning microscope using a Plan-Apochromat 63 \times /1.40 Oil DIC M27 or EC Plan-Neofluar 10 \times /0.3 M27 objective and ZEN software.

Determination of OSN coalescence. P2-IRES-tauLacZ and M72-IRES-tauLacZ mice³¹ were crossed with *Ano2^{+/-}* mice to yield *Ano2^{-/-}* and *Ano2^{+/+}* littermates that were homozygous for these reporter lines. Experiments were carried out with mice from 5 weeks to 24 weeks old. The mice were perfused with PBS followed by 4% PFA (wt/vol) in PBS. Brains were post-fixed (20 min on ice), washed in buffer A (100 mM phosphate buffer (pH 7.4), 2 mM MgCl₂, 5 mM EGTA) and incubated for 30 min in buffer A with 0.02% NP-40 (vol/vol) and 0.01% deoxycholate (wt/vol). LacZ staining was carried out overnight in buffer A with 0.2% NP-40 (vol/vol), 0.1% deoxycholate (wt/vol), 5 mM potassium ferricyanide 5 mM potassium ferrocyanide and 1 mg ml⁻¹ of X-Gal at room temperature (20–25°C). Whole-mount stained bulbs were cryoprotected in 30% sucrose (wt/vol), frozen in Tissue-Tek O.C.T. compound (Sakura) and cut into 20- μ m cryostat sections. Sections were post-stained with X-Gal and counterstained with Neutral Red.

Lysate preparation, deglycosylation and immunoblotting. Turbinates, VNO or eyes were dissected, minced and sonicated in lysis buffer (10 mM HEPES (pH 7.4), 150 mM NaCl, 5 mM EDTA, protease inhibitors, 1% SDS (wt/vol)). Lysates from brain regions and ANO1-transfected HEK cells were prepared similarly. The ANO1 expression construct encodes a 986-amino-acid human isoform (NM_018043.5). For deglycosylation, lysates were incubated with 5% β -mercaptoethanol (vol/vol) for 15 min at 55 °C, diluted 1:5 into deglycosylation buffer (50 mM HEPES (pH 7.4), 10 mM EDTA, 0.5% NP-40 (vol/vol) and protease inhibitors) and incubated overnight at 37 °C with 1 U of N-Glycosidase F (Roche Diagnostics) per 100 μ g protein. Proteins were separated on SDS-polyacrylamide gels and transferred on polyvinylidene difluoride membranes. Antibodies were used at the following dilutions: rbAno2_N3-3 antibody (1:1,000); anti-adenylate cyclase III rabbit polyclonal (1:2,000, C-20, sc-588, Santa Cruz); anti-olfactory marker protein (OMP) goat polyclonal (1:1,000, 544-10001, Wako); anti- α -tubulin mouse monoclonal (1:2,000, clone B-5-1-2, T5168, Sigma-Aldrich); anti-Cnga2 (1:100), anti-tyrosine hydroxylase (1:5,000) and anti-TMEM16A (1:250) antibodies, as in immunohistochemistry. Signals were visualized by chemiluminescence (SuperSignal West, Pierce) using a PeqLab camera system and secondary antibodies coupled to horseradish peroxidase (Chemicon). Immunoblots were repeated in three or more independent experiments.

Quantitative real-time PCR. Turbinates were dissected from *Ano2^{+/+}* and *Ano2^{-/-}* mice (three litter pairs, 25–27 weeks old) and total RNA was isolated (RNeasy Mini Kit, Qiagen). We subjected 1 μ g of RNA to DNase I (amplification grade, Invitrogen) digestion and transcribed it into cDNA using random primers and Superscript II reverse transcriptase (Invitrogen). We set up 20- μ l PCR reactions using the Power SYBR Green PCR Master Mix (Applied Biosystems) and ran the reactions in triplicates with a 40-s elongation time at 60 °C. Amplification and melting curves were monitored using a StepOnePlus Real-Time PCR System and StepOne Software (Applied Biosystems). Comparison between *Ano2^{-/-}* and wild-type littermates was carried out using the Pfaffl method and β -actin as internal control. Primers spanned introns or exon-exon boundaries and gave products of 100–200 bp. The following primer pairs were used. β -actin: 5'-TGATGATGGTGGGAATGGGTCAGAA-3', 5'-TGTGGTGCCAGATCTTCTCCATGT-3'; adenylate cyclase III: 5'-CGGCATCGAGTGTCTACGCTTC-3', 5'-GCCAGCGCTCCTTGCTGTGACTT-3'; *Ano2_{ex3-4}*: 5'-GATGTTGAACCTGGGCCACT-3', 5'-CAAGAACCTCTCTTCCCTGG-3'; *Ano2_{ex20-21}*: 5'-AGGCTGTCTCATGGAGCTGT-3', 5'-TGCTCTGGACGTTTGTAGTG-3'; *Ano6*: 5'-AAGAGGAACAGGCCCGGCCA-3', 5'-AGAAGACGGCGCTCGCACAC-3'; *Ano8*: 5'-GCGACGCGGCTGGAAGACAG-3', 5'-CGGCCAGTGGGAAGGCAGAG-3'; *Ano10*: 5'-AGCGGAAGTACTGTGCGAGGGT-3', 5'-GCAAAGGCAGCTGTAGCGG-3'; *Nkcc1*: 5'-CTGGTGGGCTGCGTTGTCTCA-3', 5'-GCGCTGTGTGGAGGATCCCC-3'.

Tissue preparation for patch-clamping. Mice were killed at 6–8 weeks by decapitation. The MOE was prepared and embedded in 4% low melting agarose (wt/vol) in normal Ringer (140 mM NaCl, 5 mM KCl, 10 mM glucose, 1 mM sodium pyruvate, 10 mM HEPES (pH 7.4), 2 mM CaCl₂, 1 mM MgCl₂; osmolality: 321 mOsm kg⁻¹). A Leica VT1200S vibratome was used to cut 200- μ m slices perpendicular to the surface. The soft tissue of the VNO was embedded⁴⁷ in 4% low melting agarose (wt/vol) and cut in 200- μ m coronal slices. Slices were kept in normal Ringer bubbled with 95% O₂/5% CO₂.

Preparation of isolated olfactory receptor neurons. The MOE was transferred to low-Ca²⁺ Ringer (140 mM NaCl, 5 mM KCl, 10mM glucose, 1 mM sodium pyruvate, 1 mM EDTA, 1 mM cysteine, 10 mM HEPES (pH 7.2)) at 4 °C. After mincing, it was digested at room temperature for 20 min with 0.2 mg ml⁻¹ trypsin in low-Ca²⁺ Ringer. The digestion was terminated with normal Ringer containing 0.2 mg ml⁻¹ BSA, 0.2 mg ml⁻¹ leupeptin and 0.025 mg ml⁻¹ DNase I. Following gentle trituration, cells were allowed to settle on poly-L-lysine-coated coverslips for 30–45 min at 4 °C.

Patch-clamp analysis. Pipettes were pulled from borosilicate glass and contained 140 mM CsCl, 4 mM HEPES (pH 7.2), 1 mM EGTA, 2 mM Mg-ATP. Free [Ca²⁺] was adjusted according to the Maxchelator program (<http://maxchelator.stanford.edu/>) to 0 μ M, 1.5 μ M and 13 μ M. OSNs from the MOE were patched

at the cell body and sometimes at the dendritic knob, with similar results. VSNS were patch-clamped at the dendritic knob. Pipette resistances were ~5 M Ω and 8–10 M Ω when patching OSNs or VSN dendritic knobs, respectively. Recordings were performed in the whole-cell configuration at room temperature in normal Ringer containing 10 mM tetraethylammonium chloride using an Axon CNS MultiClamp 700B amplifier, a Digidata 1322A interface and pClamp 10 software (Molecular Devices).

Photorelease of caged Ca²⁺ and 8-Br-cAMP. We used [6,7-Bis(carboxymethoxy)coumarin-4-yl]methyl-8-bromo-adenosine-3', 5'-cyclic monophosphate (BCMCM-caged 8-Br-cAMP), a generous gift from V. Hagen (Leibniz-Institut für Molekulare Pharmakologie) and DMNP-EDTA (Invitrogen). For uncaging 8-Br-cAMP, a UV flash lamp JML-C2 (Rapp OptoElektronik) was coupled to an inverted Zeiss Axiovert 200 microscope equipped with a 100 \times Fluor objective. Ca²⁺ was uncaged with a SP-20 UV flash lamp (Rapp OptoElektronik) coupled to an upright Olympus BX51WI microscope equipped with a 60 \times LUMPlanFL objective. Isolated OSNs were patched at room temperature in the whole-cell mode. For Ca²⁺ uncaging, the pipette solution contained 140 mM KCl, 2 mM Mg-ATP, 0.3 mM Na₂-GTP, 10 mM HEPES (pH 7.4), 3.5 mM CaCl₂ and 5 mM DMNP-EDTA. For uncaging 8-Br-cAMP, the pipette solution contained 145 mM KCl, 4 mM MgCl₂, 1 mM Mg-ATP, 0.1 mM Na₂-GTP, 10 mM HEPES (pH 7.4), 0.5 mM EGTA and 0.1 mM BCMCM-caged 8-Br-cAMP. Settings were 200 V at 1 mF and 300 V at 3 mF for uncaging 8-Br-cAMP and Ca²⁺, respectively.

Electro-olfactogram recordings. The head was cut in the sagittal plane and the endoturbinates were exposed^{11,48}. The lateral side of the head was immersed into 0.5% agarose gel with normal Ringer.

In fluid-phase EOGs the turbinates were continuously superfused with normal Ringer with or without niflumic acid. The four-barreled application pipette had two channels for continuous flow of normal Ringer, a channel for odorant-free vehicle and for odorants. The last two channels were connected to a Picospritzer (Toohey) to apply pressure steps. Mixtures of odorants, single odorant or forskolin (specified in legend to **Figure 7**, in aqueous solution containing 0.5% DMSO (vol/vol)) were used.

In the air-phase configuration, the application tube had a channel for continuous flow of humidified 95% O₂/5% CO₂. Another tube connected to a 15-ml bottle containing 2 ml of odorants or odorant-free vehicle, and a Picospritzer. Pressure steps of 200 ms were used to drive the air from the bottle to the turbinates. In both arrangements, EOGs were recorded in current clamp with an extracellular electrode filled with Ringer placed on the middle of turbinate IIb or III. EOG amplitudes from single turbinates were averaged from ten individual sweeps and subsequently averaged from several experiments.

Ion-sensitive microelectrodes. Chloride-selective electrodes were fabricated⁴⁹ from theta-glass capillaries (Warner Instruments) with tip diameters of ~3 μ m.

Ion-selective barrels were silanized with methyltrichlorosilane in dichloromethane (Fluka/Sigma-Aldrich) and backfilled with Chloride ionophore I – cocktail A' (Fluka/Sigma-Aldrich). Reference barrels contained 155 mM NaCl. To exclude artifacts by Cl⁻ diffusion, we also filled them with 40 mM NaCl and 77 mM Na₂SO₄. No differences in measured [Cl⁻] were found. Measurements with either reference barrel solution were pooled. Microelectrodes connected to a differential amplifier⁴⁹ were lowered at a low angle onto the surface of freshly dissected turbinates. [Cl⁻] was determined from calibration curves before and after measurements. Measurements were discarded when these calibrations differed by >20%.

Olfactometry. Behavioral assessment of odor detection and discrimination ability used a computer-controlled eight-channel liquid-dilution olfactometer (Knosys), as described⁵⁰. Mice were trained on a go or no-go operant conditioning task in which licking on a water delivery tube upon presentation of an S+ stimulus (odor 1) is associated with a water reward, but not after presentation of an S- stimulus (odor 2 or diluent). Mice that are able to discriminate between stimuli readily stop to answer to the S- stimulus when subjected to successive trials of S+ and S- stimuli presented in random order. Response accuracy is monitored as a measure for odor discrimination and detection ability and represents the percentage of correct answers in a block of 20 trials.

Water-deprived *Ano2*^{-/-} mice and control littermates were trained on the BEGIN program and on a simple odor discrimination task. For testing we used the D2 program with standard settings⁵⁰. Chemicals were from Sigma-Aldrich/Fluka. Mice (35–41 weeks old) were tested on 1% (-)-limonene versus 0.5% (-)/0.5% (+)-limonene (six males, two females) and on geraniol dilutions (eight males, four females) starting with the highest concentration. Trials were 200 (down to 10⁻⁵) or 400 (10⁻⁶, 10⁻⁷) and response accuracy was calculated as mean of three consecutive blocks after reaching the criterion response of 90% or of the last three blocks in 400 trials, respectively. Mice at 14 weeks old (four males, two females) were used for 1% hexanal (vol/vol) versus 1% octanal (vol/vol) discrimination, and six males and four females (at 50 weeks old) were used for the 0.4/0.6% versus 0.6/0.4% hexanal/octanal discrimination test. Three male *Cnga2*^{-/-} mice (14–16 weeks old) were tested on 1% geraniol (vol/vol).

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